FISEVIER

Contents lists available at ScienceDirect

Bioorganic & Medicinal Chemistry Letters

journal homepage: www.elsevier.com/locate/bmcl



Simple and convenient radiolabeling of proteins using a prelabeling-approach with thiol-DOTA

Carmen Wängler a,*, Ralf Schirrmacher , Peter Bartenstein b, Björn Wängler b,*

- ^a McConnell Brain Imaging Center, Montreal Neurological Institute, McGill University, University Street 3800, Montreal H3A 2B4, Quebec, Canada
- ^b University Hospital Munich, Department of Nuclear Medicine, Ludwig-Maximilians-University, Marchioninistraße 15, Munich, Germany

ARTICLE INFO

Article history: Received 19 January 2009 Revised 11 February 2009 Accepted 13 February 2009 Available online 20 February 2009

Keywords:
Radiolabeling
Radiometals
Molecular imaging
Proteins
DOTA
Thiol-maleimidecoupling

ARSTRACT

Commonly applied methods for radiometal-labeling of proteins require complex and protracted derivatization reactions of the protein and the subsequent radiolabeling is time-consuming due to the low reaction temperatures applicable. Therefore, a convenient and efficient prelabeling technique for proteins using the DOTA derivative 2,2',2"-(10-(2-(2-mercaptoethylamino)-2-oxoethyl)-1,4,7,10-tetraazacyclod-odecane-1,4,7-triyl)triacetic acid (thiol-DOTA) containing a thiol moiety for rapid and selective introduction into maleimide-derivatized proteins was developed. Thiol-DOTA was labeled with ⁶⁸Ga, ⁹⁰Y and ¹⁷⁷Lu and subsequently introduced into bovine serum albumin and a human IgG with maximum radiochemical yields of 66%. The entire radiolabeling procedure was completed after only 30 min making this a favorable new labeling technique for proteins.

© 2009 Elsevier Ltd. All rights reserved.

Although bioactive compounds labeled with radiometals as, for example, ⁶⁸Ga, ⁹⁰Y and ¹⁷⁷Lu have gained widespread interest in diagnostic imaging as well as for therapeutic purposes, only few examples of proteins labeled with radiometals can be found. Using the commonly applied radiometal chelator DOTA (1,4,7,10-tetraaz-acyclodocecane-*N*,*N*,*N*,*N*,*N*, *N*, retraacetic acid), which forms highly stable complexes with a broad variety of radiometals, high temperatures for a fast and quantitative complexation reaction are required. These conditions are suitable for the labeling of peptides and oligonucleotides but not for proteins when the chelator is conjugated to the protein prior to the radiolabeling reaction.

Recently, labeling methods for proteins with ⁶⁷Ga and ⁶⁸Ga using the chelators NOTA and HBED-CC were described. ^{1–5} However, these chelators either form stable complexes only with gallium and indium isotopes and are not applicable for the stable labeling with other important radiometals such as ⁸⁶Y, ⁹⁰Y and ¹⁷⁷Lu or are only accessible by a complex synthesis and have shown to be highly immunogenic. ^{6–8} As mentioned above, DOTA derivatives are particularly suitable for radiometal-labeling as they form very stable complexes with a broad variety of radiometal ions. Several DOTA derivatives for the modification and subsequent

E-mail addresses: carmen.waengler@med.uni-muenchen.de (C. Wängler), bjoern.waengler@med.uni-muenchen.de (B. Wängler).

labeling of proteins at mild temperatures were described for the use in post-labeling approaches as, for example, DOTA-L-p-isothiocyanato-phenylalanine, DOTA-triglycyl-L-p-isothiocyanato-phenylalanine, DOTA N-hydroxysulfosuccinimide, DOTA-maleimidoethylacetamide and active esters of DOTA. However, the derivatization reactions of the proteins are complex and time-consuming and the subsequent labeling reactions take between 30 and 60 min due to the low reaction temperature. $^{9-15}$ This is relatively long regarding the short half-life of some radionuclides such as 68 Ga ($t_{1/2}$: 68 min). In addition, the complexation reactions are mostly not quantitative due to the low reaction temperatures used to avoid protein denaturation.

Using DOTA-triglycyl-L-*p*-isothiocyanato-phenylalanine in a prelabeling approach, the labeling procedure was time-consuming and the reaction of the precomplexed nuclide with the protein required a high excess of the protein to be labeled to ensure acceptable reaction yields of the moderately reactive isothiocyanate with the protein. This resulted in a high amount of non-radioactive protein which can negatively influence the pharmacokinetic and pharmacodynamic properties of the radiolabeled compound.

Thus, a DOTA derivative allowing an efficient and rapid introduction of various radiometals into proteins without intricate and time-consuming derivatization and purification steps would be advantageous. This would also allow the use of short-lived radionuclides such as ⁶⁸Ga and should result in an uncomplicated

^{*} Corresponding authors.

synthesis of radiolabeled proteins. The DOTA derivative should furthermore allow a facile determination of the number of derivatization sites introduced per protein as strong structural changes exerted by the derivatization can easily result in a significant loss of the biological activity of the protein.^{17–19}

A DOTA derivative meeting these requirements is thiol-DOTA (3) which was synthesized according to an earlier published procedure.²⁰ This DOTA derivative, containing a thiol moiety for effective and selective introduction into maleimide-derivatized substances. showed to be oxidation-resistant in solution at pH 7.2 for several hours and also exhibits very favorable coupling properties as the Michael addition to the maleimide generally takes place within minutes. Thiol-DOTA (3) was thus investigated regarding its applicability for a prelabeling approach with ⁶⁸Ga, ⁹⁰Y and ¹⁷⁷Lu and its use as a labeling agent for the maleimide-derivatized proteins bovine serum albumin (BSA) and a human monoclonal IgG. These proteins were derivatized with Sulfo-SMCC (2) (sulfosuccinimidyl-4-(N-maleimidomethyl)-cyclohexane-1-carboxylate sodium salt) to introduce maleimide moieties and subsequently reacted with the prelabeled thiol-DOTA (5).²¹ The amount of maleimidederivatization sites per proteins was determined according to Wängler et al. by Ellman's Assay and found to be between 1.5 and 1.8.¹⁹

The prelabeling approach using thiol-DOTA (3) allowed an efficient complexation of ⁶⁸Ga, ⁹⁰Y and ¹⁷⁷Lu at high temperatures and subsequently a very rapid introduction into the maleimide-functionalized proteins under mild conditions. The final purification

step could be carried out using a size exclusion gel cartridge. Thus, this prelabeling approach enables the very rapid radiolabeling of various proteins.

A typical labeling procedure comprised the following steps (Scheme 1):

- 1. the prelabeling of thiol-DOTA (**3**, 4.35 μ g, 10 nmol) with the radiometal (230–250 MBq 68 Ga in sodium acetate buffer (1.1 mL, 0.1 M, pH 3.8) or 0.5–1.1 GBq 90 Y in sodium acetate buffer (0.5 mL, 0.4 M, pH 4.8) or 550–850 MBq 177 Lu in sodium acetate buffer (0.5 mL, 0.4 M, pH 4.8)) for 10 min at 99 °C,
- 2. the introduction of the prelabeled complex (**5**) into the maleimide-derivatized protein (**4**, 10–35 nmol)²¹ by reacting 10 min at room temperature in phosphate buffer (1 mL, 0.5 M, pH 7.2) and
- 3. the purification of the radiolabeled protein (**6**) using a NAP-10-column (GE Healthcare Bio-Sciences AB, Uppsala, Sweden).

The prelabeling of thiol-DOTA (**3**) with 68 Ga (Fig. 1A), 90 Y and 177 Lu (Fig. 1D) proceeded with high yields, giving the radiolabeled thiol-DOTA complex (**5**) in radiochemical yields of 93–98% and high radiochemical purities within 10 min at 99 °C. As the thiol-DOTA (**3**) was used in low quantities of only 10 nmol, the achieved specific activities were between 20–25 GBq/ μ mol for 68 Ga, 45–100 GBq/ μ mol for 90 Y and 50–80 GBq/ μ mol for 177 Lu.

The ⁶⁸Ga-thiol-DOTA-complex (⁶⁸Ga-**5**) (Fig. 1A) and the ⁹⁰Y-and ¹⁷⁷Lu-thiol-DOTA-complexes (¹⁷⁷Lu-**5**) (Fig. 1D) show differ-

Scheme 1. Labeling of proteins (bovine serum albumin or human IgG) with radiometals M³⁺ (⁶⁸Ga³⁺, ⁹⁰Y³⁺ or ¹⁷⁷Lu³⁺). Reagents and conditions: (i) 100 nmol protein (1), 7 equiv Sulfo-SMCC (2), 1 h, room temperature, phosphate buffer (0.5 mL, 0.1 M, pH 7.2), 95–98½²¹; (ii) 10 nmol thiol-DOTA (3), 230–250 MBq ⁶⁸Ga in sodium acetate buffer (1.1 mL, 0.1 M, pH 3.8) or 0.5–1.1 GBq ⁹⁰Y in sodium acetate buffer (0.5 mL, 0.4 M, pH 4.8) or 550–850 MBq ¹⁷⁷Lu in sodium acetate buffer (0.5 mL, 0.4 M, pH 4.8), 10 min, 99 °C; (iii) 10–35 nmol maleimide-derivatized protein (4), prelabeled thiol-DOTA from ii (5), 10 min, room temperature, phosphate buffer (1 mL, 0.5 M, pH 7.2).

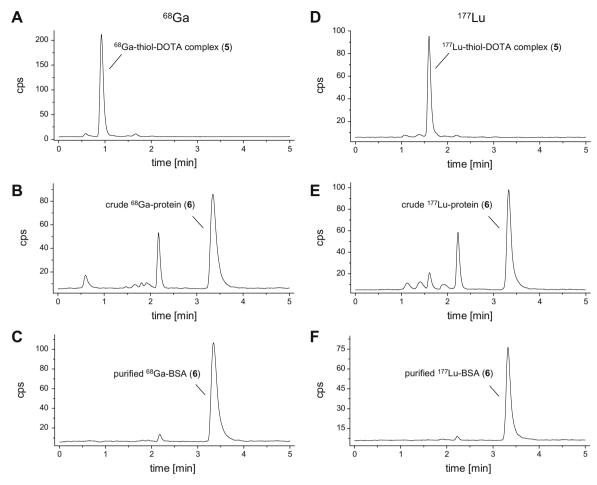


Figure 1. Analytical radio-HPLC chromatograms at different stages of the labeling reaction: A and D show the finished precomplexation reactions of thiol-DOTA with ⁶⁸Ga and ¹⁷⁷Lu, B and E the reactions of maleimide-derivatized BSA (**4**) with the labeled ⁶⁸Ga- and ¹⁷⁷Lu-thiol-DOTA complexes (**5**) and C and F the purified ⁶⁸Ga- and ¹⁷⁷Lu-labeled BSA (**6**). ⁹⁰Y labeling reactions result in equivalent chromatograms to those of ¹⁷⁷Lu.

ent retention times under the same HPLC conditions of 0.95 min and 1.6 min, respectively, which might be attributed to the different ion radius and therefore to the different complexation behavior of the radiometals. The free thiol function does not seem to contribute to the complex formation as it subsequently reacts with the maleimide-derivatized protein. Moreover, the contribution of the thiol moiety in the complexation is not likely regarding the known complex geometry of DOTA complexes.²²

In contrast to the prelabeling step yielding only the desired complexation product (**5**), its reaction with the maleimide-derivatized proteins (**4**) shows the formation of some unidentified side products (Fig. 1B and E) which are likely to consist of small labeled molecules as can be deduced from analyzing the same reaction mixtures with size exclusion FPLC.²³ However, these side products can be easily removed by the final gel filtration purification (Fig. 1C and F), yielding the pure radiolabeled proteins (**6**).

The overall radiochemical yields of the labeling reactions were between 18% and 66% and the specific activities achieved were between 1.1–3.2 GBq/ μ mol for 68 Ga, 8.1–27 GBq/ μ mol for 177 Lu and 4.5–32 GBq/ μ mol for 90 Y which is in the range of published values. 9,11,12,16

By applying Ellman's assay,²⁴ the extent of the maleimidederivatization of the proteins can be verified. Using this assay, we introduced a well defined and low number of 1.5–1.8 maleimide-derivatization sites into the proteins which results in only minor structural alteration and by this in a highly preserved biological activity of the protein to be labeled. As the maleimidederivatized proteins can be stored in solution at $4\,^{\circ}\text{C}$ for several hours without loss of maleimide reactivity, the derivatization can be accomplished before labeling, further shortening the labeling procedure.

The radiolabeling of the maleimide-derivatized proteins ($\mathbf{4}$) can be accomplished within 30 min as the prelabeling of thiol-DOTA ($\mathbf{3}$) takes only 10 min as well as the reaction of the prelabeled complex with the protein and the purification of the final product via a gel cartridge can be carried out within 5–10 min.

In summary, it has been demonstrated that thiol-DOTA can be used as a valuable tool for the convenient and rapid radiolabeling of maleimide-derivatized proteins with ⁶⁸Ga, ⁹⁰Y and ¹⁷⁷Lu. Particularly the very simple, mild and defined derivatization of the protein and the efficient radiolabeling procedure which is completed after a short time span of only 30 min make this labeling technique a valuable alternative to applied methods.

References and notes

- Eder, M.; Wängler, B.; Knackmuss, S.; LeGall, F.; Little, M.; Haberkorn, U.; Mier, W.; Eisenhut, M. Eur. J. Nucl. Med. Mol. Imaging 2008, 35, 1878.
- Wu, C.; Jagoda, E.; Brechbiel, M.; Webber, K. O.; Pastan, I.; Gansow, O.; Eckelman, W. C. Bioconjugate Chem. 1997, 8, 365.
- 3. Lee, J.; Garmestani, K.; Wu, C.; Brechbiel, M. W.; Chang, H. K.; Choi, C. W.; Gansow, O. A.; Carrasquillo, J. A.; Paik, C. H. Nucl. Med. Biol. 1997, 24, 225.
- Govindan, S. V.; Goldenberg, D. M.; Elsamra, S. E.; Griffiths, G. L.; Ong, G. L.; Brechbiel, M. W.; Burton, J.; Sgouros, G.; Mattes, M. J. J. Nucl. Med. 2000, 41, 2089.
- Ochakovskaya, R.; Osorio, L.; Goldenberg, D. M.; Mattes, M. J. Clin. Cancer Res. 2001, 7, 1505.

- 6. Cacheris, W. P.; Nickle, S. K.; Sherry, A. D. Inorg. Chem. 1987, 26, 958.
- Andereggi, G.; Arnaud-Neu, F.; Delgado, R.; Felcman, J.; Popov, K. Pure Appl. Chem. 2005, 77, 1445.
- 8. Zöller, M.; Schuhmacher, J.; Reed, J.; Maier-Borst, W.; Matzku, S. *J. Nucl. Med.* **1992**, 33, 1366.
- Grünberg, J.; Novak-Hofer, I.; Honer, M.; Zimmermann, K.; Knogler, K.; Bläuenstein, P.; Ametamey, S.; Maecke, H. R.; Schubiger, P. A. Clin. Cancer Res. 2005, 1114, 5112.
- Sabbah, E. N.; Kadouche, J.; Ellison, D.; Finucane, C.; Decaudin, D.; Mather, S. J. Nucl. Med. Biol. 2007, 343, 293.
- 11. Lewis, M. R.; Raubitschek, A.; Shively, J. E. Bioconjugate Chem. 1994, 56, 565.
- Lewis, M. R.; Kao, J. Y.; Anderson, A. L.; Shively, J. E.; Raubitschek, A. Bioconjugate Chem. 2001, 122, 320.
- Ahlgren, S.; Orlova, A.; Rosik, D.; Sandström, M.; Sjöberg, A.; Baastrup, B.; Widmark, O.; Fant, G.; Feldwisch, J.; Tolmachev, V. Bioconjugate Chem. 2008, 191, 235.
- Mier, W.; Hoffend, J.; Krämer, S.; Schuhmacher, J.; Hull, W. E.; Eisenhut, M.; Haberkorn, U. Bioconjugate Chem. 2005, 161, 237.
- Hoffend, J.; Mier, W.; Schuhmacher, J.; Schmidt, K.; Dimitrakopoulou-Strauss, A.; Strauss, L. G.; Eisenhut, M.; Kinscherf, R.; Haberkorn, U. Nucl. Med. Biol. 2005, 32, 287.
- Li, M.; Meares, C. F.; Salako, Q.; Kukis, D. L.; Zhong, G. R.; Miers, L.; DeNardo, S. J. Cancer Res. 1995, 55, 5726s.
- 17. Nikula, T. K.; Boccia, M.; Curcio, M. J.; Sgouros, G.; Ma, Y.; Finn, R. D.; Scheinberg, D. A. *Mol. Immunol.* **1995**, *32*, 865.

- Brechbiel, M. W.; Gansow, O. A.; Atcher, R. W.; Schlom, J.; Esteban, J.; Simpson, D. E.; Colcher, D. *Inorg. Chem.* 1986, 25, 2112.
- Wängler, C.; Moldenhauer, G.; Eisenhut, M.; Haberkorn, U.; Mier, W. Bioconjugate Chem. 2008, 19, 813.
- Wängler, C.; Wängler, B.; Eisenhut, M.; Haberkorn, U.; Mier, W. Bioorg. Med. Chem. 2008, 16, 2606.
- 21. Derivatization of the protein with Sulfo-SMCC: 100 nmol of the protein were dissolved in 500 μ L phosphate buffer (0.1 M, pH 7.2). To this solution was added a freshly prepared solution of 0.7 μ mol (0.31 mg) of Sulfo-SMCC (sulfosuccinimidyl-4-(N-maleimidomethyl)-cyclohexane-1-carboxylate, Sigma Aldrich) in 31 μ L H₂O/DMF 1:1 and incubated for 1 h at room temperature. The derivatized protein was purified using NAP-10 columns and phosphate buffer (0.25 M, pH 7.2) and obtained in yields of 95–98%.
- Vipond, J.; Woods, M.; Zhao, P.; Tircso, G.; Ren, J.; Bott, S. G.; Ogrin, D.; Kiefer, G. E.; Kovacs, Z.; Sherry, A. D. *Inorg. Chem.* 2007, 46, 2584.
- 23. The analytical HPLC system used was an Agilent 1200 system together with a Chromolith Performance (RP-18e, 100–4.6 mm, Merck, Germany) column. Size-exclusion FPLC was performed on a Superdex 200 10/30 GL column (Amersham Biosciences AB, Uppsala, Sweden).
- Ellman's Assay was carried out according to: Wängler, C.; Moldenhauer, G.;
 Eisenhut, M.; Haberkorn, U.; Mier, W. Bioconjugate Chem. 2008, 19, 813.
- Hermanson, G. T. In Bioconjugate Techniques; Academic Press: San Diego, 1996. Chapter 5.